



A Unified Taxonomy for Ciliary Dyneins

Erik F. Y. Hom,^{1,2} George B. Witman,³ Elizabeth H. Harris,⁴ Susan K. Dutcher,⁵ Ritsu Kamiya,⁶ David R. Mitchell,⁷ Gregory J. Pazour,⁸ Mary E. Porter,⁹ Winfield S. Sale,¹⁰ Maureen Wirschell,¹⁰ Toshiki Yagi,¹¹ and Stephen M. King^{12*}

¹Department of Molecular and Cellular Biology, Harvard University, Cambridge, Massachusetts

²FAS Center for Systems Biology, Harvard University, Cambridge, Massachusetts

³Department of Cell Biology, University of Massachusetts Medical School, Worcester, Massachusetts

⁴Department of Biology, Duke University, Durham, North Carolina

⁵Department of Genetics, Washington University School of Medicine, St. Louis, Missouri

⁶Department of Biological Sciences, Graduate School of Science, University of Tokyo, Bunkyo-ku, Tokyo, Japan

⁷Department of Cell and Developmental Biology, Upstate Medical University, Syracuse, New York

⁸Program in Molecular Medicine, University of Massachusetts Medical School, Worcester, Massachusetts

⁹Department of Genetics, Cell Biology and Development, University of Minnesota, Minneapolis, Minnesota

¹⁰Department of Cell Biology, Emory University School of Medicine, Atlanta, Georgia

¹¹Department of Cell Biology and Anatomy, Graduate School of Medicine, University of Tokyo, Hongo, Tokyo, Japan

¹²Department of Molecular, Microbial and Structural Biology, University of Connecticut Health Center, Farmington, Connecticut

Received 17 June 2011; Revised 7 September 2011; Accepted 9 September 2011

Monitoring Editor: Pekka Lappalainen

The formation and function of eukaryotic cilia/flagella require the action of a large array of dynein microtubule motor complexes. Due to genetic, biochemical, and microscopic tractability, *Chlamydomonas reinhardtii* has become the premier model system in which to dissect the role of dyneins in flagellar assembly, motility, and signaling. Currently, 54 proteins have been described as components of various *Chlamydomonas* flagellar dyneins or as factors required for their assembly in the cytoplasm and/or transport into the flagellum; orthologs of nearly all these components are present in other ciliated organisms including humans. For historical reasons, the nomenclature of these diverse dynein components and their corresponding genes, mutant alleles, and orthologs has become extraordinarily confusing. Here, we unify *Chlamydomonas* dynein gene nomenclature and establish a systematic classification scheme based on structural properties of the encoded proteins. Furthermore, we

provide detailed tabulations of the various mutant alleles and protein aliases that have been used and explicitly define the correspondence with orthologous components in other model organisms and humans.

© 2011 Wiley Periodicals, Inc.

Key Words: *Chlamydomonas*, cilia, dynein, flagella, microtubule

Introduction

The assembly and motility of eukaryotic cilia and flagella require the action of a large array of dynein microtubule motor complexes. These enzymes display distinct motile properties [Kagami and Kamiya, 1992; Moss et al., 1992a, 1992b; Sakakibara and Nakayama, 1998] and contain one or more heavy chain(s) (HCs; ~500 kDa) that exhibit ATPase and microtubule motor activity. In addition, the dynein HCs are associated with a complex array of smaller polypeptides that are necessary for motor assembly, regulation, and attachment to the appropriate cargo [reviewed in King and Kamiya, 2009]. Due to the ease of genetic and biochemical analyses, a cell architecture that allows clear observation of flagellar movement, and a sequenced genome [Merchant et al., 2007], the biflagellate green alga *Chlamydomonas reinhardtii* has become the premier model system in which to dissect the role of dyneins in axoneme-based motility and in the assembly of cilia/flagella.

Chlamydomonas expresses 16 dynein HCs that form a series of motor complexes with different functions. The outer

Additional Supporting Information may be found in the online version of this article.

Abbreviations used: DC, docking complex; HC, heavy chain; IC, intermediate chain; IFT, intraflagellar transport; LC, light chain; LIC, light intermediate chain; LRR, leucine-rich repeat; NDK, nucleoside diphosphate kinase.

*Address correspondence to: Stephen M. King, Department of Molecular, Microbial and Structural Biology, University of Connecticut Health Center, 263 Farmington Avenue, Farmington, Connecticut 06030-3305. E-mail: king@neuron.uhc.edu

Published online 22 September 2011 in Wiley Online Library (wileyonlinelibrary.com).

dynein arm, containing three distinct HCs, is required for high power output by the flagellum [Piperno and Luck, 1979; Pfister et al., 1982; Brokaw, 1999]. Two different general types of inner dynein arms, one containing a HC heterodimer and a second consisting of monomeric HC species, are needed to define the waveform [Brokaw and Kamiya, 1987; Kamiya et al., 1991] and/or for beating under high viscous load [Yagi et al., 2005]. Finally, a homodimeric dynein (here, termed the intraflagellar transport [IFT] dynein) powers retrograde IFT and is thus necessary for assembly and maintenance of the organelle [Pazour et al., 1999; Porter et al., 1999]. Although *C. reinhardtii* contains a large complement of flagellar dyneins, its genome does not encode most of the components comprising the conventional cytoplasmic dynein 1/dynactin system that in other organisms (such as mammals) is required for a wide array of microtubule-based intracellular transport activities [Pfister et al., 2006; Merchant et al., 2007; Wickstead and Gull, 2007]; the exceptions are certain light chains (LCs) employed by both conventional cytoplasmic dynein and other dynein subtypes [King et al., 1996; Harrison et al., 1998; Bowman et al., 1999].

To date, a total of fifty-four gene products have been identified in *C. reinhardtii* as integral components of these dynein motors or as factors required for their assembly in the cytoplasm, transport into the flagellum, and/or localization within the axonemal superstructure [see Cole, 2009; King and Kamiya, 2009 for reviews]. These proteins have been identified by numerous laboratories over many years utilizing a variety of methods including genetic analysis of mutants with defective flagella, direct protein biochemistry, and, more recently, comparative genomic approaches. As a result, the genes, their encoded proteins and mutant strains have been given a wide variety of names derived from various nomenclature schemes. The resulting plethora of terms and aliases has become unwieldy and complicated. Moreover, the nomenclature of the orthologous dynein components in other species is often quite distinct from that used in *C. reinhardtii*, and this continues to engender considerable confusion in the literature, and in some cases has led to the misidentification of gene products.

Historically, this general problem derives, at least in part, from the fact that many *C. reinhardtii*, sea urchin¹ and *Tetrahymena thermophila* dynein proteins were given alphanumeric assignments based on the order of their migration in SDS and/or urea polyacrylamide gels many years before any of the sequences were known. Thus, differences in migration patterns due to minor variations in size, sequence and/or charge resulted in orthologous proteins being given com-

pletely different designations. Unfortunately, the issue was compounded during annotation of the mouse and human genomes when certain dynein genes were named after their *C. reinhardtii* counterparts whereas others followed the sea urchin protein nomenclature. For example, mammalian DNAL4 was named after the LC4 component of the sea urchin outer arm dynein which is orthologous to *C. reinhardtii* LC10; confusingly, in *C. reinhardtii* LC4 denotes a calmodulin homolog and thus a member of a completely unrelated protein family. Conversely, mammalian DNAL1 was named after the *C. reinhardtii* outer arm dynein leucine-rich repeat protein LC1 (the sea urchin ortholog of which is termed LC2 in one nomenclature scheme), whereas sea urchin LC1 is a member of the Tctex1/Tctex2 protein family. This level of confusion also extends to the HCs where, for example, the gene for the 1 α HC of inner arm dynein I1/f is *DHC1* in *C. reinhardtii*, *DNAH10* in sea urchins and mammals and *DYH6* in *T. thermophila*, while the 1 β HC of that same dynein is termed *DHC10* (*C. reinhardtii*), *DNAH2* (sea urchins and mammals), and *DYH7* (*T. thermophila*).

Given the long history of these names in dynein research combined with the complexity of the gene families and the large variety of organisms involved, there seems to be no way of synthesizing a gene nomenclature/numbering scheme that is completely consistent across a broad phylogenetic spectrum and incorporates all the major model organisms while still maintaining continuity with the older literature. Consequently, as part of a re-annotation effort for the *C. reinhardtii* genome, we describe in this report a new consensus nomenclature for dynein genes in *C. reinhardtii*. Furthermore, we provide a series of tables that indicate (i) the various gene aliases, and mutant and protein names that have been used in *C. reinhardtii* and (ii) the identity of the orthologous components in a variety of other model organisms where that correspondence can be unambiguously defined.

The Nomenclature

Here, we propose new names for the *C. reinhardtii* dynein genes. The formal standard for gene names in *C. reinhardtii* is a three-letter root (all capitals) followed by a number [Dutcher and Harris, 1998]. As the dynein genes encode a wide range of protein structural and functional types, we have employed these features, as far as possible, to form the basis of the new nomenclature. A list of the proposed dynein gene roots and their derivation is provided in Table I. The assignment of new gene names, the older gene indicator(s) used in previous annotations of the *C. reinhardtii* genome, the accession number and the encoded protein products are tabulated in Table II. Whenever possible, the proposed gene names are based on previous names; e.g., *DHC1*–*DHC11* are unchanged. The nomenclature scheme also provides a rational basis for the

¹Multiple species of sea urchin have been used for biochemical studies by different laboratories depending on geographic and seasonal variables. The most commonly employed include: *Anthocidaris crassispina*, *Arbacia punctulata*, *Hemicentrotus pulcherrimus*, *Lytechinus pictus*, *Pseudocentrotus depressus*, *Strongylocentrotus droebachiensis*, *Strongylocentrotus purpuratus*, and *Triploneustes gratilla*.

Table I. Proposed Roots for *C. reinhardtii* Dynein Genes

| Gene family root | Root derivation | Characteristics of protein family |
|------------------|---|--|
| <i>DHC</i> | Dynein Heavy Chain | ATPases/motors |
| <i>DIC</i> | Dynein Intermediate Chain | WD-repeat proteins |
| <i>DLI</i> | Dynein Light Intermediate chain | Originally named based on migration between ICs and LCs. Class found only in “cytoplasmic” dyneins, including IFT dynein |
| <i>DLU</i> | Dynein components with Leucine-rich repeats | Contain $\beta\beta\alpha$ barrels derived from leucine-rich repeats |
| <i>DLX</i> | Dynein Light chain thioredoxin | Redox-sensitive thioredoxins with vicinal dithiols |
| <i>DLT</i> | Dynein Light chain Tctex1-like | Tctex1/Tctex2 family proteins; some are also found in conventional cytoplasmic dynein |
| <i>DLR</i> | Dynein Light chain Roadblock-like | Related to the roadblock light chains found in conventional cytoplasmic dynein |
| <i>DLL</i> | Dynein Light chain in LC8 family | Very highly conserved family of dimeric light chains found in many enzyme systems |
| <i>DLE</i> | Dynein Light chain in EF-hand family | Ca ²⁺ -binding components containing EF-hand motif(s) |
| <i>DCC</i> | Dynein Coiled Coil | Contain extensive regions of coiled-coil structure |
| <i>DOI</i> | Dynein Outer arm-Interacting | Associate with outer arm dynein but do not fall into other categories |
| <i>DII</i> | Dynein Inner arm-Interacting | Associate with inner arm dyneins but do not fall into other categories |
| <i>DAP</i> | Dynein Assembly PIH domain | Required for dynein assembly and contain PIH domains |
| <i>DAW</i> | Dynein Assembly WD repeat | Required for dynein assembly and contain WD-repeat motifs |
| <i>DAU</i> | Dynein Assembly leucine-rich repeat | Required for dynein assembly and contain leucine-rich repeat motifs |
| <i>DAB</i> | Dynein Assembly Blocked | Required for dynein assembly but do not fall into other categories |

naming of new genes encoding dynein subunits as these are identified; we propose these be numbered sequentially.

It is important to note that although we propose altering the gene names to yield an internally consistent scheme, we suggest that current mutant and protein names be retained so as to maintain continuity in the literature. Thus, we recommend that when describing a gene product in a publication, the corresponding gene name be used at first mention so that the gene product is unambiguously identified, and that the common protein and/or mutant names be employed thereafter. This could be readily achieved by inclusion of a brief statement such as “DHC1b (encoded at *DHC16*) is the dynein motor subunit responsible for retrograde IFT.”

Mutants, Protein Aliases, and Orthologs

Mutants defective in dynein genes have been identified through a variety of genetic screens following UV or insertional mutagenesis. These strains exhibit a range of phenotypes including various degrees of flagellar dysfunction, slow swimming, and impaired flagellar assembly depending on the mutant allele and the particular component that is altered. For example, strains unable to assemble outer dynein arms exhibit a characteristic slow, jerky swimming phenotype [Kamiya and Okamoto, 1985; Mitchell and

Rosenbaum, 1985], whereas those with defective inner arms have defects in forming bends of appropriate amplitude [Kamiya et al., 1991]. The mutant alleles that have been isolated for each component and the various aliases used for the encoded proteins are listed in Table III.

As detailed above, much confusion has built up in the literature about which dynein components are orthologous due to the long history of dynein research and the multiple naming schemes used in various organisms. Consequently, Table IV provides a listing of the current *C. reinhardtii* gene and protein names along with their orthologs (where those can be unambiguously determined) in the ciliate *T. thermophila*, the sea urchins *Anthocardaris crassispina* and *Strongylocentrotus purpuratus*, the primitive chordate *Ciona intestinalis*, the fish *Danio rerio*, and the mammal *Homo sapiens*. A more comprehensive tabulation is provided in the supplemental table available online.

In conclusion, we describe here a new consensus nomenclature for the flagellar dynein genes of *C. reinhardtii* and provide a comprehensive tabulation of the gene products and various aliases, the mutant alleles isolated for each gene, and the designations of orthologous components in other model organisms. Axonemal dyneins provide the basis for ciliary motion in all organisms with motile cilia, and IFT dynein is necessary for the assembly and maintenance of cilia in most ciliated organisms. Because of its utility for biochemical and genetic analyses, *C. reinhardtii* has been a favorite model for understanding

Table II. *C. reinhardtii* Dynein Gene Nomenclature^a

| New gene name | Original gene name | Accession number | Description of encoded protein |
|--------------------------------------|-------------------------|------------------|--|
| Heavy chains | | | |
| <i>DHC1</i> | <i>DHC1 (IDA1, PF9)</i> | Q9SMH3 | 1 α heavy chain of inner arm I1/f |
| <i>DHC2</i> | <i>DHC2</i> | XP_001694660 | Inner arm dynein species d heavy chain |
| <i>DHC3</i> | <i>DHC3</i> | XP_001696272 | Inner arm dynein heavy chain (minor species) ^b |
| <i>DHC4</i> | <i>DHC4</i> | EDP07657 | Inner arm dynein heavy chain (minor species) ^b |
| <i>DHC5</i> | <i>DHC5</i> | XP_001699742 | Inner arm dynein species b heavy chain |
| <i>DHC6</i> | <i>DHC6</i> | XP_001700741 | Inner arm dynein species a heavy chain |
| <i>DHC7</i> | <i>DHC7</i> | XP_001692695 | Inner arm dynein species g heavy chain |
| <i>DHC8</i> | <i>DHC8</i> | XP_001692092 | Inner arm dynein species e heavy chain |
| <i>DHC9</i> | <i>DHC9 (IDA9)</i> | BAE19786 | Inner arm dynein species c heavy chain |
| <i>DHC10</i> | <i>DHC10 (IDA2)</i> | Q9MBF8 | 1 β heavy chain of inner arm I1/f |
| <i>DHC11</i> | <i>DHC11</i> | XP_001694047 | Inner arm dynein heavy chain (minor species) ^b |
| <i>DHC12</i> | <i>DHC1a (PCR4)</i> | EDP05194 | Inner arm dynein heavy chain ^c |
| <i>DHC13</i> | <i>ODA11</i> | Q39610 | α outer arm heavy chain |
| <i>DHC14</i> | <i>ODA4</i> | Q39565 | β outer arm heavy chain |
| <i>DHC15</i> | <i>ODA2</i> | Q39575 | γ outer arm heavy chain |
| <i>DHC16</i> | <i>DHC1b</i> | Q9SMH5 | Dynein heavy chain that mediates retrograde IFT |
| WD-repeat intermediate chains | | | |
| <i>DIC1</i> | <i>ODA9</i> | Q39578 | IC1 from outer arm dynein |
| <i>DIC2</i> | <i>ODA6</i> | P27766 | IC2 from outer arm dynein |
| <i>DIC3</i> | <i>IDA7</i> | AAD45352 | IC140 from inner arm I1/f dynein |
| <i>DIC4</i> | <i>BOP5</i> | AAU93505 | IC138 from inner arm I1/f dynein |
| <i>DIC5</i> | <i>FAP133</i> | XM_001699649 | IFT dynein intermediate chain |
| Light intermediate chains | | | |
| <i>DLI1</i> | <i>D1bLIC</i> | AAT37069 | Light intermediate chain of IFT dynein |
| Leucine-rich repeat proteins | | | |
| <i>DLU1</i> | <i>LC1 (DLC1)</i> | AAD41040 | Outer arm dynein γ heavy chain-associated |
| <i>DLU2</i> | <i>ODA8 (MOT37)</i> | EPD09919 | ODA8 protein required for outer arm assembly |
| Thioredoxin-like light chains | | | |
| <i>DLX1</i> | <i>LC3 (DLC3)</i> | Q39592 | LC3 thioredoxin associated with outer arm β heavy chain |
| <i>DLX2</i> | <i>LC5 (DLC5)</i> | Q39591 | LC5 thioredoxin associated with outer arm α heavy chain |
| Tctex1-like light chains | | | |
| <i>DLT1</i> | <i>LC9^d</i> | AAZ95589 | LC9 present in outer arm dynein |
| <i>DLT2</i> | <i>ODA12</i> | AAB58383 | LC2 present in outer arm dynein |
| <i>DLT3</i> | <i>TCTEX1</i> | AAC18035 | Tctex1 present in inner arm I1/f |
| <i>DLT4</i> | <i>TCTEX2b</i> | DAA05278 | Tctex2b present in inner arm I1/f |
| Roadblock-like light chains | | | |
| <i>DLR1</i> | <i>ODA15 (DLC7a)</i> | AAD45881 | LC7a present in outer arm and inner arm I1/f dyneins |
| <i>DLR2</i> | <i>LC7b (DLC7b)</i> | EDP03034 | LC7b present in outer arm and inner arm I1/f dyneins |
| DYNLL/LC8 family light chains | | | |
| <i>DLL1</i> | <i>FLA14</i> | Q39580 | LC8 present in outer arm, inner arm I1/f and IFT dyneins. Also a component of the radial spokes |
| <i>DLL2</i> | <i>ODA13</i> | Q39579 | Outer arm dynein LC6 |
| <i>DLL3</i> | <i>LC10 (MOT24)</i> | EDP00562 | Outer arm dynein LC10 |

(Continued)

Table II. (Continued)

| New gene name | Original gene name | Accession number | Description of encoded protein |
|---|----------------------------|------------------|--|
| Calmodulin (EF-hand) homologs | | | |
| <i>DLE1</i> | <i>LC4 (DLC4)</i> | Q39584 | LC4 present in outer arm dynein. Binds Ca ²⁺ |
| <i>DLE2</i> | <i>VFL2</i> | P05434 | Centrin present in monomeric inner arm dyneins b, e, and g. This gene is also termed CNT1 (named for CeNTrin). Binds Ca ²⁺ |
| <i>DLE3</i> | <i>ODA14</i> | AAP49435 | DC3 component of outer arm docking complex. Binds Ca ²⁺ |
| Coiled-coil proteins | | | |
| <i>DCC1</i> | <i>ODA3</i> | AAC49732 | DC1 of the outer arm docking complex |
| <i>DCC2</i> | <i>ODA1</i> | AAK72125 | DC2 of the outer arm docking complex |
| <i>DCC3</i> | <i>ODA5</i> ^d | AAS10183 | ODA5 protein that associates with an adenylate kinase |
| Outer arm dynein interacting proteins | | | |
| <i>DOI1</i> | <i>LIS1</i> ^{d,e} | ABG33844 | LIS1 protein associates with α heavy chain of outer arm |
| Inner arm dynein interacting proteins | | | |
| <i>DII1</i> | <i>IDA4</i> | Q39604 | p28 light chain present in inner arm species a, c, and d |
| <i>DII2</i> | <i>FAP146</i> | BAG07147 | p38 associates with inner arm species d |
| <i>DII3</i> | ^d | BAF98914 | p44 associates with inner arm species d |
| <i>DII4</i> | <i>IDA5</i> | P53498 | Actin, present in inner arm dynein species a, b, c, d, e, g, and some minor species. This gene is also known as ACT1 (named for ACTin) |
| <i>DII5</i> | <i>NAP1</i> | AAC49834 | NAP, novel actin-related protein that can substitute for actin in inner arm dyneins b and g. This gene is also known as ARP12 (named for Actin Related Protein). |
| <i>DII6</i> | <i>FAP94</i> | EDP03678 | IC97 present in inner arm I1/f dynein |
| <i>DII7</i> | <i>FAP120</i> | EDP07339 | Ankyrin-repeat protein that interacts with IC138(DIC4) from inner arm I1/f |
| Dynein assembly proteins containing a PIH domain | | | |
| <i>DAP1</i> | <i>PF13 (MOT45)</i> | BAG69288 | PF13 protein required for inner/outer arm assembly in cytoplasm |
| <i>DAP2</i> | <i>IDA10 (MOT48)</i> | BAI83444 | MOT48 protein required for inner arm assembly in cytoplasm ^f |
| Dynein assembly proteins containing WD repeats | | | |
| <i>DAW1</i> | <i>ODA16</i> | AAZ77789 | ODA16 protein acts as an IFT adaptor for outer arm dynein |
| Dynein assembly proteins containing leucine-rich repeats | | | |
| <i>DAU1</i> | <i>ODA7</i> | Q09JZ4 | ODA7 is a LRR protein required for outer arm assembly in cytoplasm |
| Dynein assembly blocked | | | |
| <i>DAB1</i> | <i>PF22</i> ^g | AEC04845 | PF22 is required for assembly of outer arms |

^aAlternative gene names are indicated in parentheses in the second column.

^bYagi et al. [2009].

^cYagi (unpublished results).

^dThese genes were missing and/or not named in the *C. reinhardtii* version 3 genome catalog.

^eNot to be confused with the *LIS1* (light-influenced suppressor) locus of Dutcher et al. [1988].

^fYamamoto et al. [2010].

^gThis gene is missing from the *C. reinhardtii* version 4 genome assembly.

the composition and function of these flagellar dyneins. As research on dynein advances in *C. reinhardtii* and other model organisms with their own advantages, the nomenclature proposed here will provide a logical basis for the

naming of newly identified dynein genes and mutant alleles and facilitate comparisons between *C. reinhardtii* and the other organisms. Finally, defects in subunits of both IFT dynein and axonemal dyneins are known to result in

Table III. Nomenclature of *C. reinhardtii* Dynein Proteins and Representative Mutant Alleles

| Gene name | Mutant alleles | Protein aliases ^a |
|--------------|--|---|
| <i>DHC1</i> | <i>ida1-1</i> → <i>ida1-6</i> , <i>pf9-1</i> → <i>pf9-4</i> , <i>pf30</i> | 1α HC |
| <i>DHC2</i> | – | DHC2 |
| <i>DHC3</i> | – | DHC3 |
| <i>DHC4</i> | – | DHC4 |
| <i>DHC5</i> | – | DHC5 |
| <i>DHC6</i> | – | DHC6 |
| <i>DHC7</i> | – | DHC7 |
| <i>DHC8</i> | – | DHC8 |
| <i>DHC9</i> | <i>ida9</i> | DHC9 |
| <i>DHC10</i> | <i>ida2-1</i> → <i>ida2-6</i> | 1β HC |
| <i>DHC11</i> | – | DHC11 |
| <i>DHC12</i> | – | DHC12 |
| <i>DHC13</i> | <i>oda11</i> | α HC |
| <i>DHC14</i> | <i>oda4-1</i> → <i>oda4-4</i> , <i>oda4-s7</i> , <i>sup_{pf1-1}</i> , <i>sup_{pf1-2}</i> | β HC |
| <i>DHC15</i> | <i>oda2</i> , <i>oda2-t</i> , <i>pf28</i> , <i>sup_{pf2}</i> | γ HC |
| <i>DHC16</i> | <i>dhc1b-1</i> , <i>stf1-1</i> , <i>stf1-2</i> , <i>dhc1b-2</i> (<i>dhc1b^β</i>) | DHC1b |
| <i>DIC1</i> | <i>oda9-1</i> , <i>oda9-2</i> (V5), <i>oda9-3</i> (V8), <i>oda9-4</i> (V24), <i>oda9-5</i> (V27) | IC1 , IC78, IC80, <i>M_r</i> 78,000 |
| <i>DIC2</i> | <i>oda6-1</i> , <i>oda6-2</i> , <i>oda6-r75</i> , <i>oda6-r88</i> | IC2 , IC69, IC70, <i>M_r</i> 69,000 |
| <i>DIC3</i> | <i>ida7</i> | IC140 , <i>M_r</i> 140,000 |
| <i>DIC4</i> | <i>bop5-1</i> , <i>bop5-2</i> | IC138 , <i>M_r</i> 138,000 |
| <i>DIC5</i> | – | D1bIC , FAP133 |
| <i>DLI1</i> | <i>d1blic</i> , <i>d1blic::D1bLIC</i> (K53S), <i>d1blic::D1bLIC</i> (K53I, S54A) | D1bLIC , LIC |
| <i>DLU1</i> | – | LC1 , <i>M_r</i> 22,000 |
| <i>DLU2</i> | <i>oda8-1</i> → <i>oda8-3</i> | ODA8 |
| <i>DLX1</i> | – | LC3 , <i>M_r</i> 16,000 |
| <i>DLX2</i> | – | LC5 , <i>M_r</i> 14,000 |
| <i>DLT1</i> | – | LC9 |
| <i>DLT2</i> | <i>oda12-1</i> , ^b <i>oda12-2</i> | LC2 , <i>M_r</i> 19,000 |
| <i>DLT3</i> | – | Tctex1 |
| <i>DLT4</i> | <i>pf16</i> (D2) ^c | Tctex2b |
| <i>DLR1</i> | <i>oda15</i> | LC7a , LC7 |
| <i>DLR2</i> | – | LC7b |

(Continued)

Table III. (Continued)

| Gene name | Mutant alleles | Protein aliases ^a |
|------------------------------|--|---|
| <i>DLL1</i> | <i>fla14-1</i> , <i>fla14-2</i> | LC8 , <i>M_r</i> 8,000, 8 kDa |
| <i>DLL2</i> | <i>oda13</i> | LC6 , <i>M_r</i> 11,000 |
| <i>DLL3</i> | <i>oda12-1</i> ^b | LC10 , MOT24 |
| <i>DLE1</i> | – | LC4 , <i>M_r</i> 18,000 |
| <i>DLE2</i> (<i>CNT1</i>) | <i>vfl2-1</i> , <i>vfl2-R1</i> , <i>vfl2-R5</i> , <i>vfl2-R8</i> , <i>vfl2-R10</i> , <i>vfl2-R11</i> , <i>vfl2-R13</i> | Centrin |
| <i>DLE3</i> | <i>oda14-1</i> (V06), <i>oda14-2</i> (V16), <i>oda14-3</i> (F28), ^d <i>oda14-1::ODA14</i> (E74Q, E152Q) | DC3 |
| <i>DCC1</i> | <i>oda3-1</i> , <i>oda3-2</i> , <i>oda3-4</i> , <i>oda3-5</i> | DC1 |
| <i>DCC2</i> | <i>oda1-1</i> → <i>oda1-3</i> | DC2 |
| <i>DCC3</i> | <i>oda5-1</i> , <i>oda5-2</i> | ODA5 |
| <i>DOI1</i> | – | LIS1 |
| <i>DII1</i> | <i>ida4-1</i> → <i>ida4-3</i> | p28 |
| <i>DII2</i> | – | p38 |
| <i>DII3</i> | – | p44 |
| <i>DII4</i> (<i>ACT1</i>) | <i>ida5</i> | Actin |
| <i>DII5</i> (<i>ARP12</i>) | – | NAP |
| <i>DII6</i> | – | IC97 , IC110 |
| <i>DII7</i> | – | FAP120 |
| <i>DAP1</i> | <i>pf13-1</i> , <i>pf13-2</i> (<i>pf13A</i>), <i>pf13-3</i> | PF13 |
| <i>DAP2</i> | <i>ida10</i> , <i>mot48</i> | MOT48 |
| <i>DAW1</i> | <i>oda16</i> | ODA16 |
| <i>DAU1</i> | <i>oda7</i> | ODA7 |
| <i>DAB1</i> ^e | <i>pf22-1</i> , <i>pf22-2</i> (<i>pf22A</i>) | PF22 |

^aThe current preferred protein name is indicated first in bold type.

^bThe *DLT2* and *DLL3* genes are adjacent; both are completely deleted in *oda12-1*.

^c*pf16*(D2) lacks both the *DLT4* and *PF16* genes; the latter encodes a component of the central pair microtubule complex.

^dThe *oda14-3*(F28) allele also lacks the *RSP14* gene which encodes a component of the radial spokes.

^eThe *DAB1* gene is currently missing from the version 4 genome assembly.

human disease [Dagoneau et al., 2009; Escudier et al., 2009; Leigh et al., 2009; Merrill et al., 2009], and the homologous relationships between *C. reinhardtii* and *H. sapiens* genes clarified here should expedite identification and analysis of candidate disease genes in human patients.

Methods

The *Chlamydomonas* dynein genes identified here are the result of a *C. reinhardtii* genome reannotation initiative

Table IV. Nomenclature of Orthologous Ciliary/Flagellar Dynein Components^{a,b}

| | <i>C. reinhardtii</i> | <i>T. thermophila</i> | <i>A. crassispina</i> and <i>S. purpuratus</i> ^c | <i>Ci. intestinalis</i> | <i>D. rerio</i> | <i>H. sapiens</i> |
|-------------------------|-----------------------|-----------------------|---|--|-----------------|--------------------------------|
| | Gene | Protein | Gene (Protein) | Protein | Gene | Gene |
| Heavy chains | | | | | | |
| Inner arm I1/f | <i>DHC1</i> | 1 α HC | <i>DYH6</i> | DNAH10 | | <i>DNAH10</i> |
| | <i>DHC10</i> | 1 β HC | <i>DYH7</i> | DNAH2 | | <i>DNAH2</i> |
| Outer arm | <i>DHC13</i> | α HC | <i>DYH5</i> (γ HC) | – | | – |
| | <i>DHC14</i> | β HC | <i>DYH4</i> (β HC) | β HC (Sp-DNAH9) | | <i>DNAH9</i> |
| | | | | | | <i>DNAH11</i> |
| | | | | | | <i>DNAH17</i> |
| IFT dynein ^d | <i>DHC15</i> | γ HC | <i>DYH3</i> (α HC) | α HC (Sp-DNAH5, Sp-DNAH8, Sp-DNAH15) | | <i>DNAH5</i> , <i>DNAH8</i> |
| Inner arm group 3 | <i>DHC16</i> | DHC1b | <i>DYH2</i> | Sp-DYNC2H1 | <i>Dync2h1</i> | <i>DYNC2H1</i> |
| | <i>DHC4</i> | DHC4 | { <i>DYH8</i> | {DNAH3 | { <i>DNAH7</i> | { <i>DNAH3</i> |
| | <i>DHC5</i> | DHC5 | <i>DYH10</i> | DNAH4 | <i>DNAH12</i> } | <i>DNAH7</i> |
| | <i>DHC6</i> | DHC6 | <i>DYH12</i> | DNAH7 | | <i>DNAH12</i> |
| | <i>DHC8</i> | DHC8 | <i>DYH13</i> | DNAH12} | | <i>DNAH14</i> } |
| | <i>DHC9</i> | DHC9 | <i>DYH14</i> | | | |
| | <i>DHC11</i> | DHC11 | <i>DYH17</i> | | | |
| | | | <i>DYH18</i> | | | |
| | | | <i>DYH25</i> } | | | |
| Inner arm group 4 | <i>DHC2</i> | DHC2 | { <i>DYH9</i> | DNAH1 | | { <i>DNAH1</i> |
| | | | <i>DYH11</i> | | | <i>DNAH6</i> } |
| | | | <i>DYH16</i> | | | |
| | | | <i>DYH19</i> | | | |
| | | | <i>DYH20</i> } | | | |
| Inner arm group 5 | <i>DHC3</i> | DHC3 | { <i>DYH15</i> | – | | |
| | <i>DHC7</i> | DHC7 | <i>DYH22</i> | DNAH6 | | |
| | | | <i>DYH23</i> | | | |
| | | | <i>DYH24</i> } | | | |
| Unassigned | <i>DHC12</i> | DHC12 | | | | |

(Continued)

Table IV. (Continued)

| | <i>C. reinhardtii</i> | <i>T. thermophila</i> | <i>A. crassispina</i> and <i>S. purpuratus</i> ^c | <i>Ci. intestinalis</i> | <i>D. rerio</i> | <i>H. sapiens</i> |
|---|-----------------------------|-----------------------|---|---|--|---|
| | Gene | Protein | Gene (Protein) | Protein | Gene | Gene |
| Intermediate chains | | | | | | |
| Outer arm | – <i>DIC1</i> | – IC1 | – <i>IC2</i> | IC1 ^e IC2 (Sp-DNAI1) | IC3 ^e IC2 | TXNDC3 ^e (<i>Sptrx2</i>) <i>DNAIL1</i> |
| | <i>DIC2</i> | IC2 | <i>IC3</i> | IC3 (Sp-DNAI2) | <i>zgc:158666</i> <i>LOC100004620</i> | <i>DNAIL2</i> |
| Inner arm | <i>DIC3</i> | IC140 | <i>IC5</i> | | | <i>WDR63</i> |
| | <i>DIC4</i> | IC138 | <i>IC6</i> | | <i>Wdr78</i> | <i>WDR78</i> |
| IFT dynein | <i>DIC5</i> | D1bIC | <i>D2IC</i> | | <i>Dync2i1</i> | <i>WDR34</i> |
| Light intermediate chain^d | | | | | | |
| Light chains | | | | | | |
| Leucine-rich repeats | <i>DLU1</i> | IC1 | <i>LC1</i> | LC2 (Sp-DNAL1) | <i>zgc:92542</i> | <i>DNAL1</i> |
| | <i>DLU2</i> | ODA8 | | LRRC56 | | <i>LRRC56</i> |
| Thioredoxine-like | <i>DLXI</i> | LC3 | <i>LC3A</i> (LC3-like A) <i>LC3B</i> (LC3-like B) | – | – | – |
| | <i>DLX2</i> | LC5 | | – | – | – |
| Tctex1-like | <i>DLT1</i> | LC9 | { <i>TCT1A</i> (<i>Tctex1A</i>) <i>TCT1B</i> (<i>Tctex1B</i>)} | LC3 (Sp-DYNLT1) | – | { <i>DYNLT1</i> ^f (<i>Tctex1</i>) <i>DYNLT3</i> (<i>rp3</i>)} |
| | <i>DLT3</i> | Tctex1 | | | | { <i>TCTE3</i> |
| | <i>DLT2</i> | LC2 | { <i>LC2A</i> | LC1 (Sp-DYNLT2) | { <i>Tctex1d1</i> | <i>DYNLT3</i> (<i>rp3</i>) |
| | <i>DLT4</i> | Tctex2b | <i>LC2B</i> | | <i>Tctex1d2</i> | { <i>TCTE3</i> |
| Roadblock-like | <i>DLRI</i> | LC7a | <i>LC7A</i> | RBPH (Sp-DYNLRB1) LC7L1 (Sp-DYNLRB2) | <i>Dynlrb1</i> | <i>TCTEX1D2</i> (<i>Tctex2</i>) { <i>DYNLRB1</i> ^f |
| | <i>DLR2</i> | LC7b | <i>LC7B</i> | | | <i>DYNLRB2</i> |
| LC8 family | <i>DLL1</i> | LC8 | <i>LC8^g</i> | LC6 (Sp-DYNLL1) | { <i>Dynll1</i> <i>Dynll2</i> } | <i>DYNLRB2</i> <i>DYNLL1</i> <i>DYNLL2</i> |
| | <i>DLL2</i> | LC6 | <i>LC8x</i> (LC8-like) ^g | – | <i>zgc:100999</i> | – |
| | <i>DLL3</i> | LC10 | <i>LC10</i> | LC4 (Sp-DNAL4) | | <i>DNAL4</i> |
| EF-hand | <i>DLE1</i> | LC4 | <i>LC4A</i> <i>LC4B</i> | – | | – |
| | <i>DLE2</i> (<i>CNT1</i>) | Centrin | <i>CEN1</i> (Centrin) | | | <i>CETN1</i> , <i>CETN2</i> , <i>CETN3</i> |
| | <i>DLE3</i> | DC3 | | | | |

(Continued)

Table IV. (Continued)

| | <i>C. reinhardtii</i> | <i>T. thermophila</i> | <i>A. crassispina</i> and <i>S. purpuratus</i> ^c | <i>Ci. intestinalis</i> | <i>D. rerio</i> | <i>H. sapiens</i> |
|-------------------------|-----------------------|-----------------------|---|-------------------------|---------------------------------|---------------------------|
| | Gene | Protein | Gene (Protein) | Protein | Gene | Gene |
| Other components | | | | | | |
| Coiled-coil | <i>DCC1</i> | DC1 | | IC4 | | <i>CCDC114</i> |
| | <i>DCC2</i> | DC2 | | IC5, Axp66.0 | | <i>CCDC63</i> |
| | <i>DCC3</i> | ODA5 | | | { <i>Wdr5</i> <i>Poc1b</i> } | |
| Outer arm-interacting | <i>DOI1</i> | LIS1 | | | <i>Dnali1</i> | <i>DNALI1</i> |
| Inner arm-interacting | <i>DIII</i> | p28 | <i>p28A</i> <i>p28B</i> <i>p28C</i> | p33 (Sp-DNALI1) | | |
| | <i>DII2</i> | p38 | | ZMYND12 | | ZMYND12 |
| | <i>DII3</i> | p44 | | TTC29 | | <i>TTC28</i> |
| | <i>DII4 (ACT1)</i> | Actin | <i>ACT1 (Actin)</i> | Actin | <i>Actin</i> | <i>Actin</i> ^h |
| | <i>DII5 (ARPI)</i> | NAP | | | | |
| | <i>DII6</i> | FAP94 | | CASC1 | | <i>CASC1</i> |
| | <i>DII7</i> | FAP120 | | | | |
| Assembly factors | | | | | | |
| PIH domain | <i>DAP1</i> | PF13 | | PIH1D1 | <i>Kintouni</i> ⁱ | <i>DNAAF2</i> |
| | <i>DAP2</i> | MOT48 | | WDR69 | <i>Pih1d1</i> | <i>PIH1D1</i> |
| WD repeats | <i>DAW1</i> | ODA16 | | LRRCS50 | <i>Wdr69</i> | <i>WDR69</i> |
| Leucine-rich repeats | <i>DAU1</i> | ODA7 | | | <i>Lrrc50</i> | <i>DNAAF1 (LRRCS50)</i> |
| Blocked assembly | <i>DAB1</i> | PF22 | | | | |

^aInitial biochemical identification of proteins comprising the axonemal dyneins of various model organisms was reported by multiple groups including: for *C. reinhardtii*, Pfister et al. [1982], Piperno and Luck [1979]; for the sea urchin *Tripanestes gratilla*, Bell et al. [1979]; for *T. thermophila*, Porter and Johnson [1983]; and for *Ci. intestinalis*, Hozumi et al. [2006].

^bThis table illustrates the names of orthologous components where that can be unambiguously determined. In some cases, multiple proteins in one organism are more closely related to each other than they are to any proteins present in another organism, as indicated by brackets, { . . . } across row entries. Thus, for the monomeric inner arm HCs (and some other components), phylogenetic analysis does not provide for a clear correspondence at the level of individual proteins. However, subgroupings are more clear [see also Wickstead and Gull, 2007; Wilkes et al., 2008] and are indicated here, although it is important to note that some ambiguity still remains.

^cThere are at least two nomenclatures for sea urchin axonemal dynein components currently in use. One derives from the original protein biochemistry and early sequence analysis of outer arm dynein components performed by a number of laboratories most notably those of Ian Gibbons [e.g., Bell et al., 1979] and Kazuo Ogasawa [e.g., Ogasawa et al., 1996, and light chain sequences published only in the database]. More recent annotation of the *S. purpuratus* genome identified additional components of sea urchin dyneins and provided alternate names for some components based mainly on the scheme used in mammals [Morris et al., 2006].

^dThe IFT dynein subunits in the nematode *Caenorhabditis elegans* are known as CHE3 (heavy chain), XBx-1 (light intermediate chain), and XBx-2 (a Tctex1/Tctex2 family light chain). A dynein IC involved in IFT has not yet been unambiguously identified in *Ca. elegans*. *Ca. elegans* lacks axonemal dyneins.

^eThese ICs are modular proteins consisting of an N-terminal thio-redoxin domain followed by several catalytic nucleoside diphosphate kinase (NDK) modules. The N-terminal domain is closely related to *C. reinhardtii* LC3 (DLX1) and LC5 (DLX2). However, subunits of the *C. reinhardtii* outer arm do not contain the NDK modules.

^fMammals express two canonical Tctex1 proteins (DYNLT1 and DYNLT3) and two DYNLRB proteins. It remains uncertain which members of these groups are orthologous to the *C. reinhardtii* flagellar dynein components. Thus, both members of each group are listed.

^gThe analysis of Wilkes et al. [2007] recognized LC8 and five LC8-like sequences (ABF38951–ABF38955) in *T. thermophila*; LC8D (LC8-likeD) (ABF38954) appeared most closely related to *C. reinhardtii* LC6 (DLL2), although our analysis suggest this assignment is ambiguous with respect to other LC8-like sequences, notably LC8C (ABF38953). Furthermore, none of these *T. thermophila* LC8-like sequences contain the loop region insert that characterizes *C. reinhardtii* LC6. The most recent *T. thermophila* genome release includes only canonical LC8 and LC8E (LC8-likeE) genes. As amino acid differences occur throughout the LC8-likeA to LC8-likeE sequences, these sequences are unlikely to be generated by alternative splicing. It is possible that the current genome assembly has erroneously combined these genes into a common locus/scaffold.

^hFor organisms which express multiple actin isoforms, it has not yet been determined which isoform(s) are present in cilia/flagella.

ⁱThe kintoun (Ktu) mutant was originally identified in medaka [*Oryzias latipes*; Omran et al., 2008].

[Hom et al., in preparation], based on models generated using the gene-calling program AUGUSTUS [Stanke et al., 2008]. Proteome datasets (sources given in Supplementary Information, Table SI) for *T. thermophila* C3, *Trypanosoma brucei* TREU 927, *S. purpuratus* (sea urchin), *Ci. intestinalis* (sea squirt), *Drosophila melanogaster* (fruit fly), *D. rerio* (zebra fish), and *H. sapiens* (human) were pairwise aligned to the set of *C. reinhardtii* dyneins by context-specific BLAST [Biegert and Soeding, 2009]. Hits with bit scores within 2% of the best hit were collected and orthologs were assigned by manual inspection, mindful of the analyses by Wickstead and Gull [2007] and Wilkes et al. [2008]. Hits to multiple *C. reinhardtii* dynein genes were treated conservatively: when one-to-one ortholog associations were uncertain, homologous proteins were grouped into subclasses.

Acknowledgment

Our laboratories are supported by National Institutes of Health grants GM032843 (to S.K.D.), GM044228 (to D.R.M.), GM060992 (to G.J.P.), GM055667 (to M.E.P.), GM51173 (to W.S.S.), GM030626 (to G.B.W.), and GM051293 (to S.M.K.), and by the Robert W. Booth Endowment (to G.B.W.), a Grant-in-Aid for Scientific Research (C) from MEXT (to T.Y.), and a grant from the Ministry of Education, Culture, Sports and Technology of Japan (to R.K.). M.W. is supported by grants to W.S.S. from the National Institutes of Health (GM051173) and the National Institute on Alcohol Abuse and Alcoholism (P50-AA-13575). E.F.Y.H. was supported in part by the Jane Coffin Childs Memorial Research Fund and the NIGMS Center for Systems Biology (GM068763).

References

- Bell CW, Fronk E, Gibbons IR. 1979. Polypeptide subunits of dynein 1 from sea urchin sperm flagella. *J Supramol Struct* 11: 311–317.
- Biegert A, Soeding J. 2009. Sequence context-specific profiles for homology searching. *Proc Natl Acad Sci USA* 106: 3770–3775.
- Bowman AB, Patel-King RS, Benashski SE, McCaffery JM, Goldstein LS, King SM. 1999. *Drosophila* roadblock and *Chlamydomonas* LC7: a conserved family of dynein-associated proteins involved in axonal transport, flagellar motility, and mitosis. *J Cell Biol* 146: 165–180.
- Brokaw CJ. 1999. Computer simulation of flagellar movement. VII. Conventional but functionally different cross-bridge models for inner and outer arm dyneins can explain the effects of outer arm dynein removal. *Cell Motil Cytoskeleton* 42: 134–148.
- Brokaw CJ, Kamiya R. 1987. Bending patterns of *Chlamydomonas* flagella. IV. Mutants with defects in inner and outer dynein arms indicate differences in dynein arm function. *Cell Motil Cytoskeleton* 8: 68–75.
- Cole D. 2009. Intraflagellar transport. In: Witman GB, editor. *The Chlamydomonas Source Book*, Vol.3: Cell Motility and Behavior. San Diego: Elsevier. pp 71–113.
- Dagoneau N, Goulet M, Geneviève D, Sznajer Y, Martinovic J, Smithson S, Huber C, Baujat G, Flori E, Tecco L, Cavalcanti D, Delezoide AL, Serre V, Le Merrer M, Munnich A, Cormier-Daire V. 2009. *DYNC2H1* mutations cause asphyxiating thoracic dystrophy and short rib-polydactyly syndrome, type III. *Am J Hum Genet* 84: 706–711.
- Dutcher SK, Harris E. 1998. *Chlamydomonas reinhardtii*. Trends in Genetics Genetic Nomenclature Guide. Cambridge, UK: Elsevier Science Ltd., pp S18–S19.
- Dutcher SK, Gibbons W, Inwood WB. 1988. A genetic analysis of suppressors of the PF10 mutation in *Chlamydomonas reinhardtii*. *Genetics* 120:965–976.
- Escudier E, Duquesnoy P, Papon JF, Amselem S. 2009. Ciliary defects and genetics of primary ciliary dyskinesia. *Paediatr Respir Rev* 10:51–54.
- Harrison A, Olds-Clarke P, King SM. 1998. Identification of the *t* complex-encoded cytoplasmic dynein light chain Tctex1 in inner arm II supports the involvement of flagellar dyneins in meiotic drive. *J Cell Biol* 140: 1137–1147.
- Hozumi A, Satouh Y, Makino Y, Toda T, Ide H, Ogawa K, King SM, Inaba K. 2006. Molecular characterization of *Ciona* sperm outer arm dynein reveals multiple components related to outer arm docking complex protein 2. *Cell Motil Cytoskeleton* 63: 591–603.
- Kagami O, Kamiya R. 1992. Translocation and rotation of microtubules caused by multiple species of *Chlamydomonas* inner-arm dynein. *J Cell Sci* 103: 653–664.
- Kamiya R, Okamoto M. 1985. A mutant of *Chlamydomonas reinhardtii* that lacks the flagellar outer dynein arm but can swim. *J Cell Sci* 74: 181–191.
- Kamiya R, Kurimoto E, Muto E. 1991. Two types of *Chlamydomonas* flagellar mutants missing different components of inner-arm dynein. *J Cell Biol* 112: 441–447.
- King SM, Kamiya R. 2009. Axonemal dyneins: assembly, structure and force generation. In: Witman GB, editor. *The Chlamydomonas Source Book*, 2nd ed. Vol.3: Cell Motility and Behavior. San Diego: Elsevier. pp 131–208.
- King SM, Barbarese E, Dillman JF,III, Patel-King RS, Carson JH, Pfister KK. 1996. Brain cytoplasmic and flagellar outer arm dyneins share a highly conserved M_r 8,000 light chain. *J Biol Chem* 271: 19358–19366.
- Leigh MW, Pittman JE, Carson JL, Ferkol TW, Dell SD, Davis SD, Knowles MR, Zariwala MA. 2009. Clinical and genetic aspects of primary ciliary dyskinesia/Kartagener syndrome. *Genet Med* 11:473–487.
- Merchant SS, Prochnik SE, Vallon O, Harris EH, Karpowicz SJ, Witman GB, Terry A, Salamov A, Fritz-Laylin LK, Marechal-Drouard L, Marshall WF, Qu L-H, Nelson DR, Sanderfoot AA, Spalding MH, Kapitonov VV, Ren Q, Ferris P, Lindquist E, Shapiro H, Lucas SM, Grimwood J, Schmutz J, Cardol P, Cerutti H, Chanfreau G, Chen C-L, Cognat V, Croft MT, Dent R, Dutcher S, Fernandez E, Fukuzawa H, Gonzalez-Ballester D, Gonzalez-Halphen D, Hallmann A, Hanikenne M, Hippler M, Inwood W, Jabbari K, Kalanon M, Kuras R, Lefebvre PA, Lemaire SD, Lobanov AV, Lohr M, Manuell A, Meier I, Mets L, Mittag M, Mittelmeier T, Moroney JV, Moseley J, Napoli C, Nedelcu AM, Niyogi K, Novoselov SV, Paulsen IT, Pazour G, Purton S, Ral J-P, Riano-Pachon DM, Riekhof W, Rymarquis L, Schroda M, Stern D, Umen J, Willows R, Wilson N, Zimmer SL, Allmer J, Balk J, Bisova K, Chen C-J, Elias M, Gendler K, Hauser C, Lamb MR, Ledford H, Long JC, Minagawa J, Page MD, Pan J, Pootakham W, Roje S, Rose A, Stahlberg E, Terauchi AM, Yang P, Ball S, Bowler C, Dieckmann CL, Gladyshev VN, Green P, Jorgensen R, Mayfield S, Mueller-Roeber B, Rajamani S, Sayre RT, Brokstein P, Dubchak

- I, Goodstein D, Hornick L, Huang YW, Jhaveri J, Luo Y, Martinez D, Ngau WCA, Otilar B, Poliakov A, Porter A, Szajkowski L, Werner G, Zhou K, Grigoriev IV, Rokhsar DS, Grossman AR. 2007. The *Chlamydomonas* genome reveals the evolution of key animal and plant functions. *Science* 318: 245–250.
- Merrill AE, Merriman B, Farrington-Rock C, Camacho N, Sebald ET, Funari VA, Schibler M.J, Firestein MH, Cohn ZA, Priore MA, Thompson AK, Rimoin DL, Nelson SF, Cohn DH, Krakow D. 2009. Ciliary abnormalities due to defects in the retrograde transport protein DYNC2H1 in short-rib polydactyly syndrome. *Am J Hum Genet* 84: 542–549.
- Mitchell DR, Rosenbaum JL. 1985. A motile *Chlamydomonas* flagellar mutant that lacks outer dynein arms. *J Cell Biol* 100: 1228–1234.
- Morris RL, Hoffman MP, Obar RA, McCafferty SS, Gibbons IR, Leone AD, Cool J, Allgood EL, Musante AM, Judkins KM, Rossetti BJ, Rawson AP, Burgess DR. 2006. Analysis of cytoskeletal and motility proteins in the sea urchin genome assembly. *Dev Biol* 300: 219–237.
- Moss AG, Gatti JL, Witman GB. 1992a. The motile beta/IC1 subunit of sea urchin sperm outer arm dynein does not form a rigor bond. *J Cell Biol* 118: 1177–1188.
- Moss AG, Sale WS, Fox LA, Witman GB. 1992b. The alpha subunit of sea urchin sperm outer arm dynein mediates structural and rigor binding to microtubules. *J Cell Biol* 118: 1189–1200.
- Ogawa K, Takai H, Ogiwara A, Yokota E, Shimizu T, Inaba K, Mohri H. 1996. Is outer arm dynein intermediate chain 1 multifunctional? *Mol Biol Cell* 7: 1895–1907.
- Omran H, Kobayashi D, Olbrich H, Tsukahara T, Loges NT, Hagiwara H, Zhang Q, Leblond G, O'Toole E, Hara C, Mizuno H, Kawano H, Fliegauf M, Yagi T, Koshida S, Miyawaki A, Zentgraf H, Seithe H, Reinhardt R, Watanabe Y, Kamiya R, Mitchell DR, Takeda H. 2008. Ktu/PF13 is required for cytoplasmic preassembly of axonemal dyneins. *Nature* 456:611–616.
- Pazour GJ, Dickert BL, Witman GB. 1999. The DHC1b (DHC2) isoform of cytoplasmic dynein is required for flagellar assembly. *J Cell Biol* 144: 473–481.
- Pfister KK, Fay RB, Witman GB. 1982. Purification and polypeptide composition of dynein ATPases from *Chlamydomonas* flagella. *Cell Motil* 2: 525–547.
- Pfister KK, Shah PR, Hummerich H, Russ A, Cotton J, Annuar AA, King SM, Fisher EMC. 2006. Genetic analysis of the cytoplasmic dynein subunit families. *PLoS Genet* 2: e1.
- Piperno G, Luck DJ. 1979. Axonemal adenosine triphosphatases from flagella of *Chlamydomonas reinhardtii*. Purification of two dyneins. *J Biol Chem* 254: 3084–3090.
- Porter ME, Johnson KA. 1983. Characterization of the ATP-sensitive binding of *Tetrahymena* 30S dynein to bovine brain microtubules. *J Biol Chem* 258: 6575–6581.
- Porter ME, Bower R, Knott JA, Byrd P, Dentler W. 1999. Cytoplasmic dynein heavy chain 1b is required for flagellar assembly in *Chlamydomonas*. *Mol Biol Cell* 10: 693–712.
- Sakakibara H, Nakayama H. 1998. Translocation of microtubules caused by the $\alpha\beta$, β and γ outer arm dynein subparticles of *Chlamydomonas*. *J Cell Sci* 111: 1155–1164.
- Stanke M, Diekhans M, Baertsch R, Haussler D. 2008. Using native and syntenically mapped cDNA alignments to improve *de novo* gene finding. *Bioinformatics* 24:637–644.
- Wickstead B, Gull K. 2007. Dyneins across eukaryotes: a comparative genomic analysis. *Traffic* 8: 1708–1721.
- Wilkes DE, Rajagopalan V, Chan CWC, Kniazeva E, Wiedeman AE, Asai DJ. 2007. Dynein light chain family in *Tetrahymena thermophila*. *Cell Motil Cytoskeleton* 64: 82–96.
- Wilkes DE, Watson HE, Mitchell DR, Asai DJ. 2008. Twenty-five dyneins in *Tetrahymena*: a re-examination of the multidynein hypothesis. *Cell Motil Cytoskeleton* 65: 342–351.
- Yagi T, Minoura I, Fujiwara A, Saito R, Yasunaga T, Hirono M, Kamiya R. 2005. An axonemal dynein particularly important for flagellar movement at high viscosity: implications from a new *Chlamydomonas* mutant deficient in the dynein heavy chain gene DHC9. *J Biol Chem* 280: 41412–41420.
- Yagi T, Uematsu K, Liu Z, Kamiya R. 2009. Identification of novel dyneins that localize exclusively to the proximal portion of *Chlamydomonas flagella*. *J Cell Sci* 122:1306–1314.
- Yamamoto R, Hirono M, Kamiya R. 2010. Discrete PIH proteins function in the cytoplasmic preassembly of different subsets of axonemal dyneins. *J Cell Biol* 190: 65–71.